

# Characterization of pregnant women exposure to halogenated parabens and bisphenols through water consumption

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1 Characterisation of pregnant women exposure to halogenated parabens and bisphenols 2 through water consumption 3 Marion Albouy<sup>a,b,c,1</sup>, Yoann Deceuninck<sup>d,1</sup>, Virginie Migeot<sup>a,b,c</sup>, Manon Doumas<sup>a,c,e</sup>, Antoine 4 Dupuis<sup>a,c,f</sup>, Nicolas Venisse<sup>a,c,g</sup>, Pascale Pierre Engene<sup>a,c</sup>, Bruno Veyrand<sup>d</sup>, Thomas Geny<sup>d</sup>, Philippe 5 6 Marchand<sup>d</sup>, Bruno Le Bizec<sup>d</sup>, Emmanuelle Bichon<sup>d\*</sup>, Pascal Carato<sup>a,c\*\*</sup> <sup>a</sup> Université de Poitiers, F-86000 Poitiers, France 7 8 <sup>b</sup> Pole Biospharm Service de Santé Publique, CHU, Poitiers, France 9 <sup>c</sup> CIC INSERM 1402, UFR Médecine Pharmacie, Poitiers, France <sup>d</sup> Oniris, INRAE, LABERCA, 44307 Nantes, France 10 <sup>e</sup> IC2MP, CNRS 7285, UFR Médecine Pharmacie, Poitiers, France 11 12 <sup>f</sup> Service de Pharmacie, CHU, Poitiers, France 13 <sup>g</sup> Service de Toxicologie et Pharmacocinétique, CHU, Poitiers, France 14 15 List of e-mail addresses: 16 marion.albouy@univ-poitiers.fr; yoann.deceuninck@oniris-nantes.fr; doumasmanon@aol.com; 17 antoine.dupuis@univ-poitiers.fr; nicolas.venisse@chu-poitiers.fr; pascale.pierre.eugene@univpoitiers.fr; bruno.veyrand@oniris-nantes.fr; thomas.geny@inovalys.fr ; philippe.marchand@oniris-18 19 nantes.fr; bruno.lebizec@oniris-nantes.fr; virginie.migeot@univ-poitiers.fr; 20 emmanuelle.bichon@oniris-nantes.fr; pascal.carato@univ-poitiers.fr 21 22 \* Correspondance to: Emmanuelle Bichon, LABERCA, UMR Oniris/INRA, Route de Gachet, Site 23 de la Chanterie, CS 50707. 44307 Nantes Cedex 3. France. E-mail adress : 24 emmanuelle.bichon@oniris-nantes.fr 1

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29 ABSTRACT:

30 Exposure of pregnant women to endocrine disruptor compounds, such as parabens and bisphenol A 31 is of concern for foetal transition. Their halogenated degradation products, mainly coming from 32 water treatment plans, could be problematic as well, depending on their occurrence in drinking 33 water in the first place. Thus, 25 halogenated compounds were synthetised in order to investigate 60 34 substances (Bisphenols, parabens and their degradation products) in 325 drinking water samples coming from a French cohort study of pregnant women. Analysis was performed by tandem mass 35 36 spectrometry coupled to gas chromatography (GC-MS/MS) after SPE extraction and derivation of 37 the contaminants. Results indicate that parabens (methylparaben, n-propylparaben, ethylparaben and n-butylparaben), bisphenols S, A and F, and their degradation product, 4-hydroxybenzoic acid, 38 39 were detected up to several hundred ng/L in drinking water, with detection frequencies between 16 40 and 88%. Regarding halogenated degradation products, the highest detection frequencies were 41 found for monochlorinated products (about 50% for 2-chlorobisphenol A), which were quantified 42 up to several tens of ng/L. Such analytical approaches with broader spectrum of monitoring (i.e. 43 chemical hazards and their degradation products) constitute in the beginning of a solution to 44 exhaustively answer the questions related to the characterization of the human chemical exposome.

46	Key words:	Exposome,	Endocrine	disruptor,	drinking	water,	GC-MS/MS
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## 50 Statement of Environmental Implication

51 Parabens and Bisphenol A are become ubiquitous chimical hazards. Whereas parabens are chemical products often used in small amounts as preservatives in cosmetics, pharmaceuticals and food, 52 53 bisphenols were mainly used as basic building block for production of plastics and coatings. Their 54 toxicity was demonstrated for human health (endocrine disruption) and marine environment. They 55 could also be halogenated after waste water treatement plans. These by-products sometimes present 56 higher toxicities than their parent compounds. Therefore, this study allows to reveal for the first 57 time their occurrence in drinking water, which were sampled from pregnant women, who are both 58 at risk and vigilant.

## 60 **1** Introduction

61 Parabens and bisphenols, described as Endocrine Disrupting Chemicals (EDCs), are chemicals 62 hazards of concern. Parabens health effects are reported in vitro, e.g. on malignant ovarian [1] or 63 breast tumor [2]) or adiposity [3] and in vivo, e.g. on anogenital distance or ovary weights [4] or 64 prostate cancer [5], whereas bisphenol adverse effects are reported on bisphenol A (BPA) and 65 mainly on its S and F substitutes[6], focusing on their hormonal activity [7] their reproductive toxicities [8]. The toxicity of bisphenol and paraben degradation products or derivatives is also of 66 67 concern, with the classification of chlorinated BPA as EDCs [9]. The estrogenic activity of Cl<sub>x</sub>BPA 68 was more studied and higher than in parent compounds [10, 11]. These compounds could act as RX 69 receptor disruptors [12] but also with other nuclear receptors [10, 13, 14]. Parabens and some 70 chlorinated parabens exhibited estrogenic activities using hER competitive enzyme-linked 71 immunosorbent assay [15, 16] or a two-hybrid assay with the yeast cells transfected with hERa for 72 brominated parabens, but few studies were found for their interaction with nuclear receptors [17].

73 The internal exposure of the general population, and of pregnant women cohorts more specifically, 74 has been already demonstrated, both for parabens and bisphenols in blood and urine [18-21]. 75 Indeed, Human are mainly exposed to parabens through personal care products whereas bisphenols 76 are prevalent in numerous food and environmental matrices [22]. However, their degradation 77 products have been little studied, except as disinfection by-products [23] whereas their half-lives are 78 longer than those of their parent compounds [24, 25]. As halogenated derivatives appear during the 79 water chlorination treatment plans [12, 26-28], the presence of such halogenated derivatives in 80 drinking water is expected. Actually, these substances (bisphenols and parabens) were ubiquitous 81 found in water resources, soils, sediments, sludge. Their phenolic moiety is described as very 82 reactive with chloride and bromide radicals during water treatment plans, promoting chlorinated 83 and brominated by-products. These results have been already confirmed by pilot studies and 84 halogenated parabens were already quantified in raw sewage water [29] However, the residual 5

contribution of parent and halogenated by-products of BPA and parabens in drinking water from a
well-characterized cohort now deserves to be known. As the population most susceptible to EDCs
exposure remains the early childhood years, the characterization of the exposure of pregnant women
to these chemical hazards via drinking water seemed relevant in the first place.

To the best of our knowledge, no study reports halogenated bisphenols and parabens in the pregnancy external exposome [23, 30-32]. Therefore, we investigate 60 compounds including BPA and its substitutes BPAF, BPB, BPE, BPF, BPS, parabens, halogenated derivatives of BPA and parabens and their degradation products in 325 drinking-water samples from a French pregnant women cohort. A dedicated GC-MS/MS method was developed to better characterise the drinking water exposure of women of childbearing age.

### 95 2 Experimentals

#### 96 2.1 Pregnant women sample collection and ethical considerations

97 Pregnant French women were enrolled in the Endocrine Disruptors Deux-Sèvres (EDDS) cohort 98 study. Drinking water samples were collected during pregnancy. The EDDS cohort recruitment 99 begun in 2011 and has been described elsewhere [33]. Briefly, eligible subjects were pregnant 100 women who declared pregnancy before 14 weeks of pregnancy. Women lived in one of the 87 101 municipalities of Deux-Sèvres (France) where drinking-water was exclusively produced from 102 surface water, because EDDS cohort objectives focused on emerging chemical contaminants that 103 were more frequently found in surface water than groundwater [34, 35]. They received 1-hour visits 104 by a trained interviewer during the second and third trimester of pregnancy. Dermal water exposure 105 and ingestion frequency of tap or bottled water were particularly investigated by a questionnaires 106 expressly designed for the study. Samples of tap water were collected at home, into two 250 mL jar, 107 filled for half in the bathroom and for half in the kitchen, and at work, both at second and third 108 trimesters of pregnancy. Five samples of water treatment plan in the department were collected

during the study (10/05/2012, 10/07/2012, 8/10/2012, 13/03/2014, 25/11/2014). Informed consent was filled in by pregnant women and her partners, and the study was approved by the ethical committee with the IRD n° 2011-A00194-37.

## 112 2.2 Br<sub>x</sub>Cl<sub>y</sub>bisphenol A and Br<sub>x</sub>Cl<sub>y</sub>parabens synthesis

The synthesis of Br<sub>x</sub>Cl<sub>y</sub>BPA has been presented previously [36]. All Br<sub>x</sub>Cl<sub>y</sub>parabens were described according general synthetic routes. All chloroparaben compounds were synthesized with sulfuryl chloride and their bromo analogues with Br<sub>2</sub>. These reagents, parabens, BPA, BPAF, BPB, BPE, BPF, BPS were purchased from Sigma-Aldrich. Detailed synthesis purification procedures and characterization (<sup>1</sup>H, <sup>13</sup>C, LC-MS, CLHP) data are provided in the supplementary file n°1.

## 118 2.3 Drinking water samples

119 325 drinking water samples, corresponding to the entire set of samples originating from EDDS 120 cohort, were analyzed by the analytical strategy developed in this work. These analyses were 121 performed under reproducibility conditions since fifteen series of samples were considered over a 122 period of two months.

## 123 2.4 Solvents, reagents and commercial standard solutions

<sup>13</sup>C<sub>6</sub>-n-butyl-4-hydroxybenzoate was used as internal standard and was purchased by CIL (Tewksbury, MA,USA). 2-hydroxy-4-methoxy-benzophenone-d<sub>5</sub> was used as external standard and was purchased by CDN Isotopes (Quebec, Canada). SPE Chromabond HR-X, 0.5 g, 6 mL, were from Macherey-Nagel (Hoerdt, France). Methanol was purchased by VWR (Fontenay-sous-Bois, France), Water HPLC grade was provided from Panreac (Lyon, France) and acetonitrile was purchased by Carlo Erba (Val-de-Reuil, France). N-methyl-N(trimethylsilyl)-trifluoroacetamide (MSTFA) was provided from Fluka-Merck (Darmstadt, Germany).

#### 131 2.5 Sample preparation

Samples of 150 mL water aliquots were prepared in a polypropylene flask with 0.2 ng of internal 132 standards mixture (<sup>13</sup>C-bisphenol A, <sup>13</sup>C-bisphenol S, <sup>13</sup>C-bisphenol F, <sup>13</sup>C-2,2',6,6'-tetrachloro 133 bisphenol A, <sup>13</sup>C-2,2',6,6'-tetrabromo bisphénol A, <sup>13</sup>C-methyl-4-hydroxybenzoate, <sup>13</sup>C-butyl-4-134 hydroxybenzoate, n-heptyl-4-hydroxybenzoate-d<sub>4</sub>). Each sample was purified on a preconditioned 135 136 SPE (6 mL methanol and 6 mL water). After drying the cartridge under moderate vacuum (<15 137 mbar), analytes were eluted with 10 mL methanol / acetonitrile mixture (50:50, v/v). external 138 standard (2 ng) was added before evaporation to dryness. Derivatization was achieved by adding 20 139 µL of MSTFA reagent into the vial and heated for about 30 min at 45°C.

#### 140 2.6 GC-MS/MS analysis

141 An Agilent 7890 gas chromatograph, equipped with an Optima-17-MS column (30 m x 0.25 mm i.d., 0.25 µm film thickness) (Macherey Nagel, Hoerdt, France), was coupled to a triple quadrupole 142 143 mass analyzer Agilent 7000 (Agilent Technologies, Santa Clara, USA). The temperature program 144 was set as follows: 100°C (2 min) and 16°C min-1 until 320°C (5 min). Injector and transfer line 145 were set at 280°C and 320°C, respectively. Source temperature was set at 250°C. Electron energy was set at 70 eV. Helium was used as carrier gas at 1 mL min<sup>-1</sup>. Argon was used as collision gas at 146 147 1.5 mL min<sup>-1</sup>. Injection was performed in the pulsed splitless mode at 300°C (40 psi, 1.5 min), with 148 2 µL. Three transitions per analyte were monitored (cf. supplementary material). Data acquisition 149 and data processing were performed using the version B07.00 Mass Hunter constructor software 150 (Agilent Technologies, Santa Clara, USA). Two transitions were monitored per compound in SRM mode (cf. Table S3). 151

#### 152 2.7 QA/QC analysis

Quantification of compounds was carried out after identification (2002/657/EC criteria)[37], according to the isotopic dilution process. Calibration was checked within the concentration range [0-33.3] ng/L, nine calibration points, on the basis of RRF repeatability (RSD<20%). Method performances were estimated on the basis of the current ISO 11352:2013 standard[38], using a quality control sample spiked at 0.5 ng/150 mL and analysed 8 times all over the analysis campaign. Within-laboratory reproducibility, trueness and combined uncertainty were determined from this QC results.

160 Background contamination was checked, on the basis of the control of intra-laboratory 161 environmental contamination, considered as a major critical issue [39]. Numerous contamination 162 sources have been already highlighted for other ubiquitous contaminants, such as analytical measurement devices, SPE columns, glassware, plastic ware, solvents. Therefore, from the various 163 164 sources of contamination, the strategy applied in the present study consisted in a systematic control 165 of the environmental contamination for each compound of interest, in each batch of samples. Three 166 procedural blank samples were included in each batch of analysis, and allowed the calculation of an 167 average concentration and its relative standard deviation (RSD), if significant signals were observed 168 with a S/N>3.

Limits of detection (LOD) and quantification (LOQ) were determined with a S/N ratio of 3 on the first and the second transition, respectively. Only one consensual LOQ value was reported for each analyte, which corresponds to the highest calculated LOQ value (upperbound approach), excluding the outliers and always above the blank contamination level and the median value of the LOQs.

173 **3 Results** 

174 All the results are synthetized and reported in **Table 1**.

-	- و	Detection	LoQ	uncertainty	P50	P75	P95	66d
Compound	n quantifica	frequency	(ng/L)	c.c at ng/L)	(ng/L)	(ng/L)	(ng/L)	(ng/L)
bisphenol S	280	86%	0,7	85%	3,9	7,0	15,5	55,0
bisphenol A	95	29%	23,2	117%	<pre>&gt; O</pre>	25,8	63,0	229
bisphenol F	51	16%	0,7	59%	<pre>&gt;COQ</pre>	<loq< td=""><td>1,2</td><td>3,0</td></loq<>	1,2	3,0
bisphenol AF	7	2%	1,0	133%	<pre>&gt;</pre>	<loq< td=""><td><loq< td=""><td>1,8</td></loq<></td></loq<>	<loq< td=""><td>1,8</td></loq<>	1,8
bisphenol B	1	0%0	0,3	96%	≥100	<pre>&gt;COQ</pre>	<pre>&gt;</pre>	<pre>&gt;</pre>
bisphenol E	1	0%0	0,4	112%	<pre>&gt;</pre>	<loq< td=""><td>≥L0Q</td><td><pre>&gt;</pre></td></loq<>	≥L0Q	<pre>&gt;</pre>
2-chloro BPA	167	51%	0,4	84%	0,4	1,1	5,0	15,6
2-bromo BPA	84	26%	0,3	164%	<pre>&gt;ToG</pre>	0,3	8,3	43,2
2,6-dichloro BPA	80	25%	0,6	98%	olo1>	<loq< td=""><td>10,2</td><td>100</td></loq<>	10,2	100
2,2'-dichloro BPA	43	13%	0,8	72%	<pre>&gt;COQ</pre>	<loq< td=""><td>1,6</td><td>8,9</td></loq<>	1,6	8,9
2'-bromo-2,6-dichloro BPA	33	10%	0,3	62%	<loq< td=""><td><loq< td=""><td>0,6</td><td>2,2</td></loq<></td></loq<>	<loq< td=""><td>0,6</td><td>2,2</td></loq<>	0,6	2,2

Table 1 : First occurrence of parabens, bisphenols and their degradation products in drinking water, EDDS cohort

2-bromo-2',6-dichloro BPA	20	6%	0,4	76%	<pre>&gt;COQ</pre>	<pre>&gt;</pre>	0,4	0,9
2,2',6-trichloro BPA	17	5%	0,5	53%	<pre>&gt;COQ</pre>	<pre>&gt;</pre>	0,5	1,3
2,2'-dibromo BPA	15	5%	0,7	193%	<pre>&gt;COQ</pre>	<pre>&gt;</pre>	<pre>&gt;</pre>	83,2
2,6-dibromo BPA	15	5%	0,5	175%	<loq< td=""><td><pre>&gt;COQ</pre></td><td><pre>&gt;CLOQ</pre></td><td>8,2</td></loq<>	<pre>&gt;COQ</pre>	<pre>&gt;CLOQ</pre>	8,2
2,2',6,6'-tetrachloro BPA	14	4%	0,4	53%	<loq< td=""><td><pre>&gt;CLOQ</pre></td><td><loq< td=""><td>1,5</td></loq<></td></loq<>	<pre>&gt;CLOQ</pre>	<loq< td=""><td>1,5</td></loq<>	1,5
2-bromo-2'-chloro BPA	×	2%	0,5	59%	∂01>	<pre>&gt;COQ</pre>	<pre>&gt;COQ</pre>	1,0
2-chloro-2',6'-dibromo BPA	L	2%	0,7	171%	<pre>&gt;COQ</pre>	<pre>&gt;</pre>	<pre>&gt;</pre>	3,0
2-chloro-2',6-dibromo BPA	9	2%	0,7	171%	∂01>	<pre>&gt;COQ</pre>	<pre>dol&gt;</pre>	1,2
2,2'-dibromo-6,6'-dichloro BPA 2,6-dibromo-2',6'-dichloro BPA	+ v	2%	0,1	66%	001>	do1>	001>	0,2
2,2',6,6'-tetrabromo BPA	4	1%	0,7	57%	∂01>	<pre>&gt;COQ</pre>	<pre>&gt;COQ</pre>	0,7
2-bromo-6-chloro BPA	4	1%	0,4	60%	<pre>&gt;COQ</pre>	<pre>&gt;</pre>	 COQ>	0,6
2-chloro-2',6,6'-tribromo BPA	4	1%	0,3	72%	∂01>	<pre>&gt;</pre>	 COQ	0,3
2,2',6-tribromo BPA	ε	1%	1,3	126%	<loq< td=""><td><loq< td=""><td><loq< td=""><td><pre>&gt;</pre></td></loq<></td></loq<></td></loq<>	<loq< td=""><td><loq< td=""><td><pre>&gt;</pre></td></loq<></td></loq<>	<loq< td=""><td><pre>&gt;</pre></td></loq<>	<pre>&gt;</pre>
2-bromo-2',6,6'-trichloro BPA	1	0%0	0,4	59%	<loq< td=""><td><pre>&gt;</pre></td><td><loq< td=""><td><pre>&gt;COQ</pre></td></loq<></td></loq<>	<pre>&gt;</pre>	<loq< td=""><td><pre>&gt;COQ</pre></td></loq<>	<pre>&gt;COQ</pre>

ethyl 3-chloro-paraben	119	37%	0,6	39%	<pre>&gt;COQ</pre>	0,8	1,7	2,6
methyl 3-bromo-paraben	54	17%	0,1	18%	<l0q< td=""><td><pre>&gt;COQ</pre></td><td>0,4</td><td>1,9</td></l0q<>	<pre>&gt;COQ</pre>	0,4	1,9
methyl 3-bromo-5-chloro-paraben	41	13%	0,5	54%	<loq< td=""><td><loq< td=""><td>2,1</td><td>4,4</td></loq<></td></loq<>	<loq< td=""><td>2,1</td><td>4,4</td></loq<>	2,1	4,4
methyl 3,5-dichloro-paraben	39	12%	0,7	26%	<pre>&gt;</pre>	<loq< td=""><td>2,3</td><td>7,9</td></loq<>	2,3	7,9
methyl 3-chloro-paraben	16	5%	0,9	32%	<loq< td=""><td><loq< td=""><td><pre>&gt;</pre></td><td>1,9</td></loq<></td></loq<>	<loq< td=""><td><pre>&gt;</pre></td><td>1,9</td></loq<>	<pre>&gt;</pre>	1,9
methyl 3,5-dibromo-paraben	10	3%	6,8	42%	<pre>&gt;COQ</pre>	<pre>&gt;COQ</pre>	≥L0Q	12,8
butyl 3,5-dichloro-paraben	7	2%	0,4	35%	<loq< td=""><td><loq< td=""><td><pre>&gt;</pre></td><td>0,8</td></loq<></td></loq<>	<loq< td=""><td><pre>&gt;</pre></td><td>0,8</td></loq<>	<pre>&gt;</pre>	0,8
propyl 3-bromo-paraben	9	2%	0,3	68%	<pre>&gt;COQ</pre>	<pre>&gt;COQ</pre>	≥L0Q	0,4
propyl 3,5-dibromo-paraben	4	1%	1,4	41%	<loq< td=""><td><pre>&gt;ToQ</pre></td><td><pre>&gt;</pre></td><td>2,1</td></loq<>	<pre>&gt;ToQ</pre>	<pre>&gt;</pre>	2,1
butyl 3,5-dibromo-paraben	c	1%	0,9	77%	<pre>&gt;COQ</pre>	<loq< td=""><td><pre>&gt;COQ</pre></td><td><loq< td=""></loq<></td></loq<>	<pre>&gt;COQ</pre>	<loq< td=""></loq<>
ethyl 3,5-dichloro-paraben	ŝ	1%	0,3	24%	<loq< td=""><td><pre>&gt;TOG</pre></td><td><pre>&gt;COQ</pre></td><td><loq< td=""></loq<></td></loq<>	<pre>&gt;TOG</pre>	<pre>&gt;COQ</pre>	<loq< td=""></loq<>
butyl 3-bromo-paraben	7	1%	0,3	27%	<loq< td=""><td><pre>&gt;COQ</pre></td><td>≥L0Q</td><td><loq< td=""></loq<></td></loq<>	<pre>&gt;COQ</pre>	≥L0Q	<loq< td=""></loq<>
ethyl 3,5-dibromo-paraben	7	1%	1,4	52%	<l0q< td=""><td><pre>&gt;TOG</pre></td><td><pre>&gt;TOG</pre></td><td><loq< td=""></loq<></td></l0q<>	<pre>&gt;TOG</pre>	<pre>&gt;TOG</pre>	<loq< td=""></loq<>
ethyl 3-bromo-paraben	2	1%	0,5	18%	<l0q< td=""><td><pre>&gt;COQ</pre></td><td><pre>&gt;COQ</pre></td><td><loq< td=""></loq<></td></l0q<>	<pre>&gt;COQ</pre>	<pre>&gt;COQ</pre>	<loq< td=""></loq<>
propyl 3,5-dichloro-paraben	5	1%	0,4	92%	<loq< td=""><td><loq< td=""><td><loq< td=""><td><loq< td=""></loq<></td></loq<></td></loq<></td></loq<>	<loq< td=""><td><loq< td=""><td><loq< td=""></loq<></td></loq<></td></loq<>	<loq< td=""><td><loq< td=""></loq<></td></loq<>	<loq< td=""></loq<>

propyl 3-chloro-paraben	2	1%	0,3	22%	<pre>&gt;COQ</pre>	<pre>&gt;</pre>	<pre>&gt;Too</pre>	oo1>
butyl 3-chloro-paraben	-	0%0	0,3	35%	<loq< td=""><td><pre>&gt;</pre></td><td><pre>&gt;Too</pre></td><td><pre>&gt; O</pre></td></loq<>	<pre>&gt;</pre>	<pre>&gt;Too</pre>	<pre>&gt; O</pre>
propyl 3-bromo-5-chloro-paraben	1	0%0	0,3	37%	<pre>&gt;COQ</pre>	oo1>	 COQ	<loq< td=""></loq<>
butyl 3-bromo-5-chloro-paraben	0	0%0	1,0	40%	<pre>&gt;COQ</pre>	olo_1>	 COQ	<pre>&gt;ToG</pre>
ethyl 3-bromo-5-chloro-paraben	0	0%0	1,4	44%	<pre>&gt;COQ</pre>	oo1>	 COQ	<pre>&gt; To</pre>
4-hydroxybenzoic acid	135	42%	24,7	174%	<loq< td=""><td>55,1</td><td>114</td><td>259</td></loq<>	55,1	114	259
3-chloro 4-hydroxybenzoic acid	128	39%	0,9	95%	<loq< td=""><td>1,6</td><td>4,0</td><td>5,2</td></loq<>	1,6	4,0	5,2
3,5-dibromo-4-hydroxybenzoic acid	92	28%	2,8	120%	<loq< td=""><td>3,8</td><td>29,9</td><td>50,5</td></loq<>	3,8	29,9	50,5
3,5-dichloro 4-hydroxybenzoic acid	76	23%	1,4	134%	<loq< td=""><td><pre>&gt;</pre></td><td>11,7</td><td>24,0</td></loq<>	<pre>&gt;</pre>	11,7	24,0
3-bromo-4-hydroxybenzoic acid	47	14%	0,8	87%	<loq< td=""><td><pre>&gt;</pre></td><td>3,0</td><td>7,2</td></loq<>	<pre>&gt;</pre>	3,0	7,2
methyl paraben	285	88%	6,0	22%	19,0	32,6	81,7	362
n-propyl paraben	280	86%	1,5	12%	7,7	14,4	48,7	136
ethyl paraben	271	83%	1,3	27%	5,0	9,7	19,9	36,5
n-butyl paraben	207	64%	1,0	16%	1,5	3,1	10,8	15,3
iso-butyl paraben	11	3%	1,0	18%	<pre>&gt;COQ</pre>	<pre>&gt;</pre>	<pre>&gt;Tod</pre>	2,7

benzyl paraben	7	2%	0,6	138%	<loq< th=""><th><l0q< th=""><th><pre>&gt;COQ</pre></th><th>1,0</th></l0q<></th></loq<>	<l0q< th=""><th><pre>&gt;COQ</pre></th><th>1,0</th></l0q<>	<pre>&gt;COQ</pre>	1,0
n-pentyl paraben	4	1%	1,3	66%	<pre>&gt;COQ</pre>	<pre>&gt;COQ</pre>	<pre>&gt;COQ</pre>	1,4
iso-propyl paraben	0	%0	1,0	21%	<pre>&gt;COQ</pre>	<pre>&gt;COQ</pre>	<pre>&gt;TOG</pre>	<loq< td=""></loq<>
n-heptyl paraben	0	%0	0,9	55%	≥	<pre>&gt;COQ</pre>	<pre>&gt;COQ</pre>	<pre>COQ</pre>

#### 176 3.1 Performances of the analytical method

177 The GC-MS/MS acquisition method was assessed enough selective to provide confident results 178 regarding the correct analyte identification. An extracted ion chromatogram is given in Figure S1 to illustrate the separation quality of halogenated parabens which have never been shown before. The 179 180 sensitivity of the analytical method was considered fit for purpose for our study, with LOOs 181 generally below the ng/L concentration level, as reported by Valbonesi et al. [40], except for BPA 182 and 4-hydroxybenzoic acid (cf. Table S1). Actually, their well-known contamination background was detected in all samples [41, 42], coming from the environmental contamination, which 183 artificially increased the LOQ value (around [20-25] ng L<sup>-1</sup>). To the best of our knowledge, this 184 185 issue of BPA contamination is not well reported in the literature either because LOQ of other 186 methods are higher the background contamination (LOQ=500 ng/L, [43]) or using the signal to 187 noise ratio of 3 to determine a technical LOQ without taking into account the environmental 188 contamination, as determined in the study of Valbonesi et al. [40] for example, with LOQ close to 189 the ng/L. In the implemented experimental conditions, no background contamination was observed 190 for the other compounds throughout the analysis campaign, neither in analytical measurement 191 devices nor in the laboratory environment, demonstrating the non-ubiquitous status of these 192 compounds. LOQs of halogenated compounds are also similar to those found by Fan et al. [44], 193 with levels below ng/L on chlorinated BPA in drinking water.

The resulting extended uncertainties, expressed within a 95% confidence range, were determined in the range of 24% for n-propyl-4-paraben and 375% for 2,2',6-trichloro BPA, for which no labelled internal standard was used (cf. Table S1), with the worst uncertainties reported for brominated bisphenols and compounds with highest LOQ (close or above 1 ng/L). Recoveries yields are also given for information in Table S2. They are included in the range [96-114] % depending on the compound monitored.

#### 201 3.2 General occurrence of contaminants in drinking water samples

Four compounds were high frequently detected (above 66 %): bisphenol S and methyl/ethyl/npropyl-parabens; 5 compounds were regularly detected (within 33-66%): 2-dichloroBPA, ethylchloro/nbutyl-parabens, 3-chlorohydroxybenzoic acid and hydroxybenzoic acid and 47 were detected at lower frequencies. Four compounds were never detected: butyl/ethyll 3-bromo-5-chloroparabens and iso-propyl/n-heptyl-parabens.

### 207 3.3 Occurrence of bisphenols and parabens in drinking water samples

The detection frequency of BPS and methyl-, n-propyl- and ethyl-parabens are above 80% while nbutyl-paraben appears in 60 % and BPA in 29% of the samples.

The medians of the water concentrations are in the range of [1.5-19] ng L<sup>-1</sup>. The highest concentrations are found methylparaben > n-propylparaben > ethylparaben > bisphenol S > nbutylparaben. Maximal concentrations (at the 99<sup>th</sup> percentile) were above 100 ng L<sup>-1</sup> for methylparaben > BPA > n-propylparaben.

BPA presents the highest measured concentrations within the bisphenol group whereas bisphenol S is found much more frequently, probably thanks to its lower LOQ, at significant level (at P50: from  $3.9 \text{ ng L}^{-1}$  to 55 ng L<sup>-1</sup>).

217 Methyl-, propyl-, ethyl- and butyl-parabens were very frequently detected, with consistent 218 concentrations, particularly for methyl- and n-propylparaben. The 4-hydroxybenzoic acid, which 219 represents the main degradation product of parabens, was also frequently detected (42 %) at level 220 up to several hundred of ng  $L^{-1}$ .

#### 221 3.4 Quantification of Br<sub>x</sub>Cl<sub>y</sub> parabens in 325 water samples

222 The most frequently detected compound was ethyl-3-chloroparaben which was clearly detected and quantified in 37% of the analyzed samples. Their corresponding 75<sup>th</sup> percentile and 99<sup>th</sup> percentile 223 values were 0.8 and 2.6 ng L<sup>-1</sup> respectively, with a maximal concentration at 18.6 ng/L. Indeed, the 224 225 sensitivity of the method has allowed the systematic detection of ethyl-3-chloroparaben at ppt level whereas the ethyl-3-bromoparaben was much less detected (in only 2 % of samples). Then, methyl-226 227 Br<sub>x</sub>Cl<sub>y</sub>-parabens were detected more frequently (in the [5-20] % of detection frequency range) and in higher concentrations, up to several ng  $L^{-1}$ , than other halogenated parabens, i.e. ethyl, propyl and 228 butyl ones. 229

Propyl- and butyl-Br<sub>x</sub>Cl<sub>y</sub>-parabens were significantly less frequently detected than methyl- and
ethyl- ones, with detection frequencies ranges below 2 %, depending on the halogen substitution.
However, when concentrations were measurable, they could reached several ng/L in drinking water,
like for propyl-3,5-dibromoparaben with 99<sup>th</sup> percentile at 2.1 ng L<sup>-1</sup>.

## 234 3.5 Quantification of Br<sub>x</sub>Cl<sub>y</sub> bisphenols in 325 water samples

235 The most frequently detected and identified compound was 2-chloroBPA with a detection frequency of 51 % and a concentration range of [0.4-16] ng/L (until the 99<sup>th</sup> percentile). In parallel 236 2-bromoBPA was quantified in 26 % of the samples studied on a concentration range of [0.3-43] 237 ng/L (until the 99<sup>th</sup> percentile). Both dichloroBPAs (2,2'-dichloroBPA and 2,6-dichloroBPA) were 238 239 also detected with significant detection frequencies of 13 % and 25 % and until 9 ng L<sup>-1</sup> and 100 ng 240  $L^{-1}$ , respectively. Mixed chlorinated and brominated molecules were also unambiguously identified with lower frequencies, from 1/325 to 10 % for 2-bromo-2',6,6'-trichloro BPA and 2'-bromo-2,6-241 dichloro BPA, respectively. 242

243 3.6 Quantification of Br<sub>x</sub>Cl<sub>y</sub>-hydroxybenzoic acid in the 325 drinking water samples

The halogenated degradation products of 4-hydroxybenzoic acid were also monitored, based on their mono- and di- Br/Cl substitution. The monochlorinated compound was the more frequent degradation product found (39 % of detection frequency) followed by the dibrominated one (28 %), the dichlorinated one (23 %) and the monobrominated one (14 %). The highest concentrations were found for the dihalogenated forms (Br<sub>2</sub> or Cl<sub>2</sub>) where drinking water concentrations reached up to several tens of ng L<sup>-1</sup>, whereas the monohalogenated forms stay below 10 ng L<sup>-1</sup>.

250 4 Discussion

#### 251 **4.1** Uncertainty measurement

252 Whereas several studies has already reported endocrine disruptor concentrations in drinking water 253 [40, 45, 46], uncertainty measurement remain poorly described in the associated articles. Indeed, recoveries and accuracy has been often preferred than uncertainties in old studies, to assess the 254 255 quantification performances, like for BPA [42] or for chlorinated parabens [28]. Indeed, Li et al. 256 [41] have developed a method focused on BPA quantification in water report accuracy values 257 determined on spiking levels well above ng/L, i.e. in the range of [300-20000] ng/L. At such high concentrations, BPA uncertainties were described at  $\pm$  40 % in the range of [500-2000] ng/L, far 258 259 away the environmental contamination level and the level of interest in drinking water.

In our study, uncertainties could reach several hundred percent, notely for degradation products. As labelled halogenoparabens and labelled of mix chloro- and bromo-bisphenols were not available as analytical standards, uncertainty measurement was led with non-mimetic internal standard for several compounds. Consequently, several extended uncertainties appeared very high. Nevertheless, the knowledge about the performances of this developed method firstly allows to assess the frequency of detection of halogenated bisphenols and parabens in drinking water. Then, each quantification value is always given with its associated uncertainty, according to the expectation of
the ISO 17025:2017 standard [43]

268

## 269 4.2 Native compounds of bisphenols and parabens

The significant occurrence of bisphenols and parabens in the drinking water samples assessed in this work, particularly those of bisphenol S and methyl/n-propyl/ethyl parabens, confirms the great interest to monitor these native substances in drinking water.

This study reveals first higher BPS detection frequency and the highest concentrations for BPA in 273 274 drinking water in the 325 collected drinking water samples, BPS detection frequencies of 86 % found in this study are considerably higher than those found in India [47] (29 %) or in China [48] 275 276 where BPS has been never detected, whereas BPA detection frequencies found in this study (29 %) 277 are below those found in the two last papers, i.e. 40 %. The other BPs analogues are more rarely 278 found in the drinking water samples, despite a detection frequency of 16 % for BPF, which was not 279 yet reported at this level in other studies. BPA concentrations still the highest quantified BP in the 280 samples of this study (P99=229 ng/L) compared to BPs analogues. The concentration range 281 quantified in the present study is in accordance with that reported by that reported in drinking water 282 samples from China (4.7–512 ng/L) [44], France (2–16.9 ng/L [45] and 28 ng/L [49]), and South 283 Africa (23 ng/L [49]). The prevalence of BPS regarding BPA in this study could be explained both 284 by the highest LOQ found for BPA, censoring the lowest contamination levels of BPA, and the 285 prohibition of BPA use in favor of BPS or BPF (which only reaches 19 % of detection frequency) when the sampling was made. The polarity of BPS (logP = 1.9), higher than the one of BPA 286 287 (logP=3.3), could also play a part in its higher contamination of drinking water. The occurrence of 288 each bisphenol seems to be rather linked to their technical use, the highest concentrations being 289 observed for BPA, and a lower but frequent detection for BPS.

290 Concerning parabens, this present study reports equivalent detection frequencies that those reported 291 in Vietnam [50] (around [80-100] %) for methyl-, ethyl-, propyl- and butylparabens but much 292 higher detection frequencies than that reported in Egypt [46], or in Spain and in Morocco [51], which did not exceed 20 % of detection for methyl-, propyl- and butylparaben. Concentrations in 293 294 methylparaben determined in this study are similar to the results found in Brazil in 2018, in the 295 range of [90-242] ng/L [52], but higher than the concentrations found in the Mediterranean basin, 296 *i.e.* around 50 ng/L in Egypt in 2020 [46], [17-40] ng/L in Spain in 2015 [53] or in Vietnam in 297 2022[50]. Whereas the drinking water concentrations of n-propylparaben (P99=136 ng/L) were 298 higher than that reported by Radwan et al. [46], the n-butylparaben concentrations are closed 299 between both studies.

300 Parabens were usually used as preservatives in cosmetics and pharmaceuticals, but also in food 301 commodities and industrial products. Their widespread production and use has led to their aqueous 302 environment leaking. These products were ubiquitous found in water resources, either from 303 wastewater treatment discharges or as a runoff from atmosphere particles [54]. The occurrence and 304 the concentration of parabens in drinking water samples seems to be also linked to their polarity, the 305 highest polar paraben (methylparaben) being detected in more concentrations and in more samples 306 than the less polar ones. However, methyl- and ethyl-parabens are still the prevalent parabens in the 307 environment [53]. This observation can also explain their prevalence in these drinking water 308 samples.

#### 309 4.3 Halogenated parabens

Ethyl-3-chloroparaben and methyl- $Br_xCl_y$ -parabens were the most commonly found halogenated parabens in the drinking water samples. As mentioned above, their high occurrence is explained by the fact that their precursors (methyl and ethyl-parabens) being the most ones present in the environment [53]. Ethylchloroparaben was the most frequently detected halogenated by-products,

314 i.e. quantified in 37 % of the drinking water samples in the range of [0.6-2.6] ng/L whereas halogenated methylparabens were found with lower detection frequencies (from 3 to 17 %). Higher 315 316 concentrations were reported for dihalogenated than monohalogenated methylparabens, with highest concentrations for dibrominated (up to 12.8 ng/L), then for dichlorinated (from 0.7 to 7.9 317 318 ng/L) and for bromochloro-methylparaben (from 0.5 to 4.4 ng/L). Higher occurrence of 319 dihalogenated compared to monohalogenated parabens have already been reported in wastewater and sludge samples [28] but never in drinking water samples. The concentration of Br<sub>x</sub>Cl<sub>y</sub>paraben 320 321 obviously depends to the initial concentration of the native paraben, to the bromide concentration in 322 water and to the potential chlorination treatment, simultaneously. In this study, we have observed 323 that these three conditions were often united, and this work has demonstrated the great interest to 324 monitor not only parabens but also halogenoparabens for the next Human exposure studies.

#### 325 4.4 Halogenated bisphenols

326 Halogenated bisphenols were commonly found in the drinking water samples of this study, 327 particularly 2-chlorobisphenol A, 2-bromobisphenol A and both dichloroBPA (2,2'-dichloroBPA 328 and 2,6-dichloroBPA), whereas they has not been detected ten years ago in drinking water sample 329 from the same location (only 8 drinking water samples) [45]. Nevertheless, the same chlorinated 330 compounds are commonly found in tap water (n=21) of patient underwent a breast surgery [36], but 331 with higher detection frequency and concentrations in this present study for mono- and 332 dichlorinated compounds. In details, quantitative results were given in our study and in the study of 333 Dumas et al. [36] at 51% vs 24% for monochloroBPA, 13-25% vs 9% for dichloroBPA, 5% vs 9% 334 for trichloroBPA, 4% vs 33% for tetrachloroBPA, respectively. This last compound could come 335 from another source because of its use as flame retardant, which could lead to an overestimation of its concentration in their studied samples. The chlorinated BPA detection frequencies found in this 336 337 study are still much lower than those found in China by Fan et al. in 2013 [44]. Actually, mono-

and dichloroBPA has been almost systematically detected in this last study, while tri- and 338 339 tetrachloroBPA were detected in more than half of the Chinese drinking water samples. The 340 chlorinated BPA concentrations found in the present study were similar to those reported in the 341 study of Fan et al., except for dichloroBPA, and particularly for the 2,6-dichloroBPA isomer, for 342 which concentrations were significantly higher ([0,6-100] ng/L) in this study, whereas they didn't 343 exceed 6 ng/L in the Fan study [44]. Finally, all the halogenated BPA degradation products were detected at least once in all the 325 samples of the present study. Once again, the simultaneous 344 345 monitoring of all these halogenated forms in the same analysis is of great interest to better 346 characterize the Human exposure to pollutants. Moreover, recent toxicological studies on halogenated BPA demonstrated an ER $\alpha$  reduced activation, and significant impacts on PPAR $\gamma$  and 347 348 inflammatory response (IL6 and IL8 expression) for selected halogenated BPA species [55], which 349 confirms the interest of following all halogenated compounds simultaneously.

## 350 4.5 Halogenated hydroxybenzoic acid degradation products

351 The monochlorinated compound is the most frequently degradation product detected. Terasaki et al. 352 already published concentrations of dichlorohydroxybenzoic acid and monochlorohydroxybenzoic acid in river water from Japan at [12-150] ng/L and [11-74] ng/L, respectively [56], which are in the 353 354 same order of concentration as the present study The detection frequency of brominated by-355 products were found equivalent to the chlorinated ones, at concentrations up to 50 ng/L for 356 dibromohydroxybenzoic acid. This result has never been published before. As hydroxybenzoic acid, its halogenated forms could come from other chemicals and cannot be directly linked to only the 357 358 paraben degradation pathway in drinking water. Actually, 3,5-dibromo-4-hydroxybenzoic acid has 359 already been described as a soil degradation product of benzonitrile herbicides such as bromoxynil 360 and ioxynil [57]. Therefore, the monitoring of such halogenated hydroxybenzoic acid compounds 361 could cover several origins, performing a quick overview of several chemical families 362 simultaneously. At that time, these compounds can be easily monitored with the multiresidue363 method.

#### 364 4.6 Strengths and limits

365 The 60 compounds studied present quite different physicochemical properties, which lead to 366 questionable quantitative performances:

- Indeed, the greater the number of compounds to be monitored, the more difficult it is to
   maintain excellent quantitative performance. However, additional internal standards
   mimetic of halogenated bisphenols and parabens should improve the measurement
   quality.
- The environmental contamination management should be improved in order to reduce LOQ at their minimum for all the ubiquitous compounds which can come from different sources at different concentrations through the lab. For each substance, source contamination assessment and reduction are a huge but relevant work to investigate contaminants at ultra-trace levels.
- 376 This multiresidue method allows the monitoring of a wide range of BPA by-products but does not cover others halogenated BPA substitutes degradation products. That would be 377 very relevant to cover a wide range of potential chemicals since native compounds (BPS 378 379 an F particularly) are found in the study samples. Using a time of flight mass 380 spectrometer would be able to increase the number of analytes monitored 381 simultaneously, paying attention to the LOQs which may not perform as well as in QqQ. 382 This also shows that classical analytical approaches of the targeted type (i.e. with a 383 priori) considerably limit knowledge of the reality of the exposome and therefore 384 constitute a major obstacle to the study of possible causal links between exposure to 385 certain chemical substances and certain pathologies suspected of being associated. The

386 development of analytical approaches without a priori (e.g. suspect screening and non-387 targeted screening) constitute in this sense the beginning of a solution to exhaustively answer the questions related to the characterization of the human chemical exposome. 388 389 It should be considered at this stage to characterize the toxicity of the hazards revealed in 390 our study. The use of New Approach Methodologies (NAM) based on (Q)SAR models 391 (e.g., Leadscope, DEREK), Read-across (e.g., OECD QSAR Toolbox), in vitro High 392 Throughput Screening (e.g., ToxCast, Tox21) or HTS assays (bioactivity evaluation at 393 the molecular or cellular level) could allow to approach the type of toxicity of these 394 chlorinated decontamination products relatively quickly. With the availability of exposure data via drinking water consumption, this better knowledge of the hazards 395 396 should make it possible to better characterise the associated risk and, if necessary, to be 397 able to enlighten the various stakeholders, e.g. drinking water producers, health 398 authorities, the population including women of childbearing age.

## 399 **5.** Conclusion

400 Sixty native and halogenate derivatives of BPA and parabens have been reported in drinking water 401 collected from pregnant women, with significant detection frequencies for monochlorinated BPA, 402 methylparaben and ethylparaben; and detection of a numerous halogenated degradation products 403 over the 44 monitored in the 325 drinking water samples included in this study. Thanks to a 404 dedicated analytical strategy especially developed for the study, using GC-MS/MS measurement, 405 and the synthesis of purified standards, our study demonstrates the great interest to monitor 406 halogenated bisphenols and parabens in drinking water samples to improve the external exposure 407 knowledge of pregnant women. Most of recent studies have focused on BPA and, to a lesser extend 408 parabens, or other EDs (such as triclosan, benzophenone, dichlorophenol) [58] but no one deal with 409 their halogenated derivatives to the best of our knowledge. In parallel, recent toxicological data 410 reinforce the interest of continuing to measure the Human exposure of these halogenated 24

411 contaminants[55]. Based on these first results, extended EDC list is available to work on the412 association between the pregnant women exposure and birth outcomes.

413

## 414 *Authors contributions*

415 PC and BLB contributed to, and were responsible for, initiating, coordinating and conducting the

416 project and studies. MD contributed to the preparation of the halogenated BPA and Parabens,

417 coordinated by PC. MAL and VM are principal investigators of the EDDS Cohort. MAL, AD, NV,

418 PPE, VM contributed to collecting drinking water samples during pregnancy. BLB contributed to

419 coordinating the analytic development. YD, EB, BV, TG and PM contributed to developing the

420 method for the identification and quantification of the 60 targeted molecules on gas chromatograph

421 coupled to a triple quadrupole mass analyzer, YD and EB contributed to data interpretation. MAL,

422 YD, EB and PC contributed to manuscript preparation. EB and PC contributed to submitting the

- 423 manuscript.
- 424 <u>Declaration of Interest</u>
- 425 The authors declare that they have no known competing financial interests or personal relationships
- 426 that could have appeared to influence the work reported in this paper.
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